



DESIGN AND IN VITRO EVALUATION OF MUCOADHESIVE MINITABLETS FOR NASAL DRUG DELIVERY OF SUMATRIPTAN

SHIVA KUMAR YELLANKI^{*1}, RAJKAMAL BIGALA¹, JAWAD ALI SYED², SAMBIT KUMAR DEB³, SHARADA GORANTI⁴

¹Ganga Pharmacy College, Jawaharlal Nehru Technological University, Nizamabad, India. E-mail: shiva_kmr1984@yahoo.com

²Department of Chemistry, Umea University, Umea -90734, Sweden.

³College of Pharmacy, SRIPMS, MGR Medical University, Coimbatore, Tamilnadu, India.

⁴Sri Padmavathi Mahila Vishwa Vidyalay, Thirupathi, Andra Pradesh, India.

ABSTRACT

The aim of the present investigation was to develop Sumatriptan Nasal Mucoadhesive Minitablets using different mucoadhesive polymers Chitosan, Carbopol 971P and Gum copal along with Methocel. Mucoadhesive minitables of Sumatriptan were prepared by direct compression method. The minitables were evaluated for thickness, hardness, swelling index, mucoadhesion and in vitro drug release. All postcompressional parameters were found to be within acceptable standard limits. It was observed that mucoadhesive minitables contained polymer blend of Carbopol, Carbopol/Chitosan, Carbopol/ Gum copal were successfully controlled the release of drug up to 7 days. The release data were fit into different kinetic models (zero order, first order and Korsmeyer- Peppas powers law equation) the mechanism of drug release was found to follow zero order release. N1 and N6 formulations were selected as optimum formulations based on evaluation studies

Key words: Nasal drug delivery, Sumatriptan succinate, Minitablets.

INTRODUCTION

The nasal route of delivery has been used for delivery of drugs for treatment of local diseases such as nasal allergy, nasal congestion and nasal infections. Recent years have shown that the nasal route can be exploited for the systemic delivery of drugs such as low molecular weight polar drugs, peptides and proteins that are not easily administered via other routes than by injection, or where a rapid onset of action is required. From the pharmacokinetic standpoint, absorption is rapid, which provides a faster onset of action compared to oral and intramuscular administration. Hepatic first-pass metabolism is also avoided, allowing increased, reliable bioavailability^{1,2}.

Sumatriptan is an agonist for a vascular 5-hydroxytryptamine receptor subtype (probably a member of the 5-HT_{1D} family). The vascular 5-HT₁-receptor subtype that Sumatriptan activates is on human basilar artery and in the vasculature of human dura mater and mediates vasoconstriction. This action in humans correlates with the relief of migraine headache. Sumatriptan has previously been shown to have a low oral bioavailability in human volunteers (15%). The intranasal route may be a viable alternative for self administration where this limitation could be overcome. To overcome the rapid clearance and to facilitate the absorption, nasal mucoadhesive materials are used in nasal formulations³.

In context of the above principles, a strong need was recognized for the development of nasal mucoadhesive dosage form to deliver Sumatriptan (model drug) to increase the efficiency of the drug, providing control release. The aim of the present study was to design and evaluate nasal mucoadhesive minitables of Sumatriptan that would increase its residence time in the nasal cavity and at the same time increase the absorption of drug and hence its bioavailability.

MATERIALS AND METHODS

Materials

Sumatriptan Succinate was supplied by Natco Pharma Ltd, Hyderabad. Chitosan was provided by India sea foods, Cochin. Methocel K4M, Carbopol 971P and Talc were received as gift samples from FDC Limited, Mumbai. Gum copal received as gift sample from Genuine chemical co. Mumbai. All other reagents and chemicals used were of analytical reagent grade.

Methods

Preparation of Mucoadhesive Minitablets^{4,5}

The powder mixture Sumatriptan succinate (2% w/w), Carbopol 971P (90 & 45 % w/w), Gum copal (90 & 45 % w/w), Chitosan (90 & 45 % w/w), Methocel K4M (5 % w/w), and Talc (3 % w/w), was obtained by homogeneously mixing the different compounds in a pestle and a mortar. The minitables (diameter 3 mm, 10 mg) were prepared by compressing the powder mixture at a force of 1.000 kN using an eccentric compression machine (Korsch Type EKO; Berlin, Germany). The production method was based on direct compression. Table 1 depicts the composition of all formulations.

Table 1: Composition of nasal mucoadhesive minitables

Ingredients	Formulations (% w/w)					
	N1	N2	N3	N4	N5	N6
Carbopol 971 P	90	-	-	45	-	45
Gum copal	-	90	-	45	45	-
Chitosan	-	-	90	-	45	45
HPMC K4M	5	5	5	5	5	5
Talc	3	3	3	3	3	3
Sumatriptan Succinate	2	2	2	2	2	2

Evaluation of mucoadhesive minitables

Content uniformity^{6,7}

Ten Minitablets were weighed and triturated to get fine powder. Weight equivalent to 2% w/w of Sumatriptan Succinate was dissolved in 25 ml of phosphate buffer pH 6.2 and sonicated for 10 min. 1 ml of this solution was withdrawn and drug content was determined by using UV spectrophotometer at 265nm after suitable dilution. The experiments were performed in triplicate, and average values were reported.

Swelling study^{8,9}

Mucoadhesive Minitablets were weighed individually (W_0) and placed separately in petri dish containing 50 ml of phosphate buffer pH 6.2. The Petri dishes were placed in an incubator maintained at $37 \pm 0.5^\circ\text{C}$. The tablets containing hydrophilic polymers swell in the medium. At 4th h, the tablets were removed from the Petri dish, tablet was then reweighed (W_t) and the % swelling index were calculated using the following formula.

$$\% \text{ WU} = \frac{(W_t - W_0)}{W_0} \times 100$$

Where, WU - Water uptake

W_t - Weight of tablet at time t

W_0 - Weight of tablet before immersion

Ex vivo mucoadhesion strength ¹⁰

In the present study, bovine cheek pouch was used as a model mucosal surface for bioadhesion testing. The bovine cheek pouch was procured from slaughter house, then excised and trimmed evenly from the sides. It was then washed in phosphate buffer (pH 6.2) and was preserved in the same or used immediately. Detachment force method was used to study the ex vivo mucoadhesion of tablets. The modified balance method was used to assess the tendency of mucoadhesive material to adhere to mucosal membrane. The left pan was replaced with a Teflon block B ring hung by a number of metallic rings. The 15mm diameter bovine cheek mucosa was attached to Teflon block A, and the tablet was attached to Teflon B using an adhesive. Block B and was lowered on block A kept in jacketed glass beaker filled with test medium (40ml of phosphate buffer pH 6.2 at 37°C). The right pan of the balance was replaced with a light weight beaker. By keeping suitable weight on the right hand side the pans were balanced so that Teflon block B attached with the tablet rest on the membrane attached to block A. After contact time of 4 min, weight was increased in the beaker on the right-hand pan by adding water until the tablet detached from the membrane. The excess weight in mg to the right hand side gave the mucoadhesive strength of the tablet.

In vitro nasal diffusion study ¹¹

In vitro nasal diffusion study was done by using diffusion cell (Figure 1). A glass cylinder with both ends open, 10 cm height, 3.7 cm outer diameter and 3.1 cm inner diameter was used. Nasal mucosa of sheep was separated from sub layer bony tissue and stored in distilled water containing few drops of gentamycin sulphate injection. After the complete removal of blood from mucosal surface it was tied to one end of donor compartment.

One Minitablet was taken in one cell (donor compartment) and fixed on mucosal surface and the cell was immersed in a beaker containing 40 ml of phosphate buffer (receptor compartment) of pH 6.2 were used for study. The cell was immersed to a depth of 1cm below the surface of phosphate buffer in the receptor compartment, agitated by a magnetic stirrer and temperature maintained at 37±1°C throughout the study. Aliquots of 1ml were withdrawn periodically at intervals of 1 d for a period of 7 d and each time equal volume was replaced with fresh phosphate buffer previously heated to 37±1°C. The amount of drug release was estimated using UV spectrophotometer at 265 nm after suitable dilution.

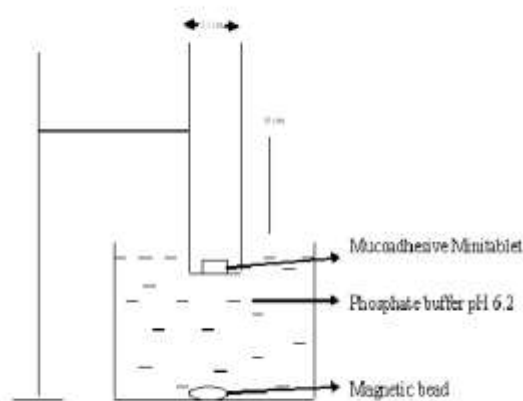


Figure 1: In vitro nasal diffusion study

RESULTS AND DISCUSSION

N1 and N6 formulations were selected as optimum formulations based on evaluation studies. These formulations are showing good mucoadhesion properties due to presence of Carbopol and Chitosan mucoadhesive polymers. Mucoadhesion studies reveal that formulations containing carbopol showed higher mucoadhesion property, due to which the formulation was retained for a longer duration on nasal mucosa.

Carbopol, Chitosan and Gum copal were selected as matrix forming and mucoadhesive polymers which help in mucoadhesion of minitabkets and slow release of the drug. HPMC is included in all formulations for maintaining the viscous nature of all formulations after absorbing the medium. Tablets were found to be satisfactory when evaluated for thickness (1.9±0.122 mm); Hardness (5.02±0.136 kg/cm²), Friability less than 1% and Drug content (98.2% – 99.9%).

Swelling index describes the amount of water that is contained within the hydrogel at equilibrium and is a function of the network structure, hydrophilicity and ionization of functional groups. Results of swelling index are shown in Table 2. From the results it can be concluded that swelling increases with time because polymer gradually absorbs water due to its hydrophilicity. The outermost layer of the polymer hydrates, swells and a gel barrier is formed at the outer surface. In the present study, all formulations had same concentrations of polymer. The swelling index was highest for tablets of formulation N2 (166.15±2.12 %) and least N1 (90.0±1.25 %). This indicates that Gum copal stores more water content in matrix than Chitosan and Carbopol.

Table 2: Drug content, swelling index and mucoadhesion strength of nasal mucoadhesive minitabkets

Formulations	Drug content (%) Mean±(SD)	Swelling index (%) Mean±(SD)	Mucoadhesion Strength (g) Mean ± (SD)
N1	99.9±0.12	90.0±1.25	24±2.12
N2	98.75±0.14	166.15±2.12	15.14±1.4
N3	98.2±0.85	144.04±1.04	19.78±2.4
N4	98.5±1.2	122±1.9	21.5±1.9
N5	99.7±1.3	161.4±2.4	17.04±1.75
N6	99.05±1.6	109.18±1.4	22.85±2.78

The ex vivo mucoadhesion strength for all formulations were determined after 4 min from contact times (Table 2). Tablets containing higher proportion of Carbopol showed higher mucoadhesion than Chitosan and Gum copal. The strength of mucoadhesion of all formulations were found in the order of N1 > N6 > N4 > N3 > N5 > N2 (Figure 2).

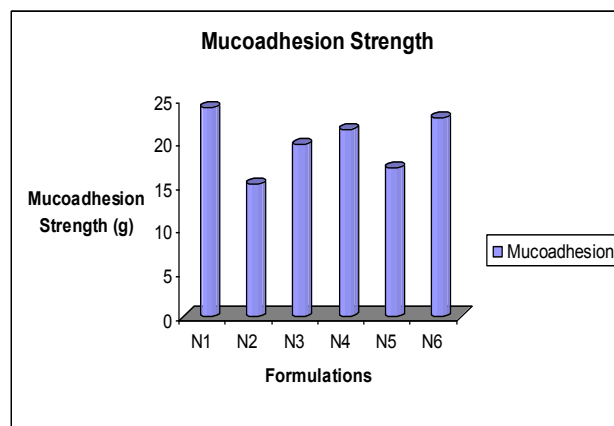


Figure 2: Mucoadhesion strength (g) of nasal mucoadhesive minitabkets

In-vitro release studies were carried out by diffusion process. The result revealed (Table 3) that release rate of the Sumatriptan Succinate was controlled for a period of time with N1 (i.e. with 90% Carbopol). The plots of the cumulative % drug release against time in days were found to be almost linear. Figure 3 showed the in vitro drug release plots of the formulations. This indicates release of the drug was controlled by diffusion controlled mechanism.

CONCLUSION

In present study, it is concluded that Sumatriptan Succinate Nasal Mucoadhesive Minitabkets can be successfully prepared using mucoadhesive polymers. The postcompressional parameters of Mucoadhesive Minitabkets (hardness, thickness and drug content)

Table 3: *In vitro* cumulative % drug release of nasal mucoadhesive minitables

Formulations	% Cumulative drug release						
	Day 1	Day 2	Day 3	Day 4	Day 5	Day 5	Day 6
N1	33.45	52.85	62.85	67.04	71.95	74.45	76.48
N2	39.57	62.78	74.85	86.45	94.12	97.85	99.72
N3	41.75	64.5	83.75	88.75	93.79	95.5	97.95
N4	36.78	57.45	66.85	73.95	78.25	81.02	84.23
N5	46.74	71.45	84.75	91.85	94.96	97.21	98.98
N6	34.52	55.42	63.75	66.78	73.89	77.45	81.75

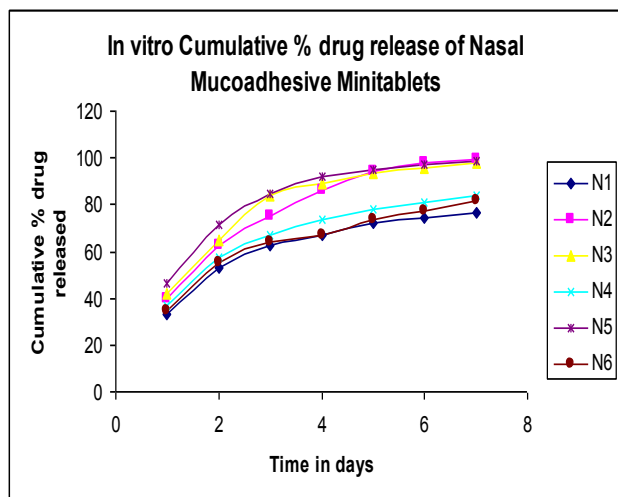


Figure 3: *In vitro* cumulative % drug released

were within acceptable official IP limits. Results of mucoadhesion test indicated that carbopol polymer increases mucoadhesion properties of Minitables. *In vitro* release results indicated that the drug release was more controlled in carbopol formulations.

The present study was a satisfactory preliminary attempt in development of Mucoadhesive intranasal Minitables of Sumatriptan Succinate.

REFERENCES

1. Lisbeth I. Nasal drug delivery-Possibilities, Problems and Solutions. *J. Cont. Release.* 2003; 87:187-198.
2. Lim ST, Forbes B, Berry DJ, Martin GP, Brown MB. *In vivo* evaluation of hyaluronic/chitosan micro particulate delivery

systems for the nasal delivery of gentamicin in rabbits. *Int. J. Pharm.* 2002; 231:73-82.

3. Umrethia ML, Majithia RJ, Majithia JB, Ghosh PK, Murthy RSR. Sumatriptan in plasma and brain tissue. *Ars. Pharm.* 2006; 47(2): 199-210.
4. Choonara YE, Pillay V, Carmichael T, Danckwerts MP. Studies on a Novel Doughnut-Shaped Minitablet for Intraocular Drug Delivery. *AAPS PharmSciTech.*2007; 8(4): Article 118.
5. Wim W, An V, Marijke M, Dhondt M, Els Ad, Philippe K, Jean PR, Annick L. Ocular Bioerodible Minitables as Strategy for the Management of Microbial Keratitis. *Investigative Ophthalmology and Visual Science.* 2004; 45: 3229-3233.
6. Weyenberg W, Vermeire A, Remon JP, Ludwig A. Characterization and *in vivo* evaluation ocular bioadhesive minitables compressed at different forces. *J. of Contr. Rel.* 2003; 89: 329-340.
7. Weyenberg W, Vermeire A, Haese ED, Vanhaelewyn G, Kestelyn P, Callens F, Nelis HJ, Remon JP, Ludwig A. Effect of different sterilization methods on the properties of bioadhesive powders and ocular minitables, and clinical evaluation. *Eur. J. Pharm. Sci.* 2004; 23: 77-87.
8. Patel VM, Prajapati BG, Patel HV, Patel KM. Mucoadhesive Bilayer Tablets of Propranolol Hydrochloride. *AAPS Pharm Sci Tech.* 2007; 8(3): 1-12.
9. Deshpande AA, Shah NH, Rhodes CT, Malick W. Development of a novel controlled release system for gastric retention. *Pharm. Research.* 1997; 14(6): 815-819.
10. Majithiya RJ, Raval AJ, Urnrethia ML, Gosh PK, Murthy RSR. Enhancement of Mucoadhesion by Blending Anionic, Cationic and Nonionic Polymers. *Drug Delivery Technology.* 2008 Feb.
11. Gupta GD, Gaud RS. Release rate of Nimesulide from different gallants. *Ind. J. Pharm. Sci.* 1999; 61 (4):227-230.